

STEREOCHEMICAL MICELLAR KINETIC DIFFERENTIATION: FACTS OR ARTIFACTS?

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The study of micelle catalyzed organic reactions^{1,2} and, in particular, micellar stereochemical processes,³ due to the analogy postulated between enzymes and micelles,⁴ has become an area of increasing interest. The recent observation of Bunton *et al.*⁵ that (R)- and (S)-*p*-nitrophenyl O-methylmandelates (I) are saponified in borate buffers in the presence of cetyltrimethyl ammonium bromide (CTABr) at identical rates while the (R/S) mixture is saponified at a rate greater than either the (R)- or (S)-enantiomer is thus of extreme importance in its implications on the nature of micellar catalysis.

Possible chiral substrate-micelle interactions are summarized in the Table. Enantiomeric substrates must interact and/or react with achiral micelles in enantiomeric modes, *i.e.*, at kinetically identical rates. The interaction and/or reaction of enantiomeric substrates with chiral micelles or of diastereomeric substrates with either achiral or chiral micelles must occur in diastereomeric modes, *i.e.*, at kinetically non-identical rates.⁶

TABLE

Chiral Substrate-Micelle Interactions

<u>Substrates/Micelle</u>	<u>Achiral</u>	<u>Chiral</u>
enantiomers	enantiomeric	diastereomeric
diastereomers	diastereomeric	diastereomeric

The observations of Bunton *et al.*⁵ are readily explicable within this framework: since (R)- and (S)-I react at identical rates in the CTABr system they obey the enantiomeric substrate-achiral micelle relationship, indicating no possible micellar chirality on the reaction time scale. The non-identical rates for racemic and optically pure ((R) or (S)) I are encompassed only by the diastereomeric substrate-achiral micelle relationship, *i.e.*, aggregation of substrate molecules (RR or SS *vrs.* RS) on a finite kinetic time scale. Such lengthy interactions are heretofore unprecedented, although on the nmr time scale they have been observed.⁷

Reinvestigation of the saponification of (R/S)- and (R)-I in sodium tetraborate decahydrate buffer (0.01 molar) and CTABr ($0.5\text{--}8.0 \times 10^{-3}$ molar) or CTACl ($1.0\text{--}8.0 \times 10^{-3}$ molar) indicates that the rates of saponification of both the racemic and optically pure esters proceed, within experimental error, at identical rates ($k_{\psi}^{\max} 1.4 \times 10^{-1} \text{ sec}^{-1}$ at 3×10^{-3} molar CTABr at one month and $1.1 \times 10^{-1} \text{ sec}^{-1}$ at 4×10^{-3} molar CTACl at two months (*vide infra*)).⁸ At other concentrations of micelle similar results are obtained ((CTABr) $\times 10^3$ molar, $k_{\psi} \times 10^2$ (R ester) sec^{-1} , $k_{\psi} \times 10^2$ (RS ester) sec^{-1} ; at one month: 1.0, 11.1 ± 0.3 , 10.9 ± 0.4 ; 4.0, 11.4 ± 0.2 , 11.8 ± 0.8 ; at ten months: 0.5, 2.0 ± 0.1 , 2.1 ± 0.1 ; 4.0, 5.3 ± 0.4 , 5.0 ± 0.5). (Errors of the orders of magnitude reported herein correspond to errors in the determination of $t_{\frac{1}{2}}$ of less than one second or errors in the determination of A_t of *ca.* 0.001 absorbancy unit.) Thus, the previous claim for stereochemical differentiation and the suggestion⁵ that chiral substrate interactions must be considered in this case is not substantiated.

A possible explanation for the previously reported⁵ stereochemical differentiation was discovered during the course of this work: the individual values of k_{ψ} at fixed micelle concentrations are dependent on the age of the micelle-buffer solution. Thus, k_{ψ} at 1×10^{-3} molar CTABr is $10.9 \pm 0.4 \times 10^{-2} \text{ sec}^{-1}$ after one month and $2.0 \pm 0.1 \times 10^{-2} \text{ sec}^{-1}$ after ten

months. The corresponding values at 4×10^{-3} molar CTABr are $11.4 \pm 0.2 \times 10^{-2} \text{ sec}^{-1}$ and $5.31 \pm 0.42 \times 10^{-2} \text{ sec}^{-1}$ at 1 and 10 months, respectively. Corresponding runs on freshly recrystallized active and racemic esters were all performed within a matter of days in the present work.

We are currently investigating the nature of this time dependency in micelle catalyzed (CTABr and CTACl) saponification reactions; preliminary indications are that neither carbon dioxide uptake nor microbial degradation of micelle solutions is responsible for the observed time dependency.

FOOTNOTES AND REFERENCES

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8. Reactions performed on a Gilford Model 2400S thermostatted at $25.00 \pm 0.05^\circ$ at 400 nm ($p\text{-NO}_2\text{C}_6\text{H}_4\text{O}^-$ appearance). Final substrate concentration 1.4×10^{-5} molar in 0.5 vol % purified p -dioxane. (R/S)-I, mp $48\text{-}49^\circ$,

lit:⁵ 46-48°; (R)-I, mp 54-55°, $[\alpha]_D^{25}$ -111.7 ± 1.6° (c 1.5, 95% ethanol),
lit:⁵ 49-50°; CTABr, mp 239-241° dec., lit:⁹ 227-235° dec., CTACl, mp
232-233° dec., lit:¹⁰ 240°. Each kinetic run performed a minimum of three
times, correlation coefficients for individual runs 0.99 or greater.

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